

Photosynthetic characteristics of phytoplankton off Adélie Land, Antarctica, during the austral summer

Toru Hirawake*, Hiroo Satoh, Takashi Ishimaru and Yukuya Yamaguchi

*Department of Ocean Science, Tokyo University of Fisheries,
Konan 4-chome, Minato-ku, Tokyo 108-8477*

Abstract: The photosynthesis-irradiance characteristics (P-E curves) and quantum yields of natural phytoplankton were investigated in the Southern Ocean off Adélie Land, Antarctica, during the austral summer. Data were acquired at eight stations during a cruise of T/V Umitaka-Maru III. The photosynthetic P-E curves showed low light adaptation of phytoplankton. Mean value (\pm standard deviation) of the P-E curve parameters, α^* , and I_k , were $0.014 (\pm 0.013) \text{ mgC (mg chl.}a\text{)}^{-1} \text{ h}^{-1}$ ($\mu\text{mol photons m}^{-2} \text{ s}^{-1}\text{)}^{-1}$ and $76 (\pm 55) \mu\text{mol photons m}^{-2} \text{ s}^{-1}$, respectively. Although phytoplankton were adapted to low irradiance, the phytoplankton in the SCM were not fully adapted to the low irradiance prevailing at those depths. P^*_{max} in the studied region was low (mean of $0.66 (\pm 0.37) \text{ mgC (mg chl.}a\text{)}^{-1} \text{ h}^{-1}$) and generally lower than the previously reported values in waters near the Antarctic Peninsula. The maximum quantum yield varied widely, ranging from 0.001 to 0.038 mol C (mol photons absorbed) $^{-1}$ at the surface and from 0.007 to 0.092 mol C (mol photons absorbed) $^{-1}$ near the bottom of the euphotic zone. These values were within the range of published data. Comparison of photosynthetic parameters with historical data indicated that primary productivity from remotely sensed data for the whole of the Southern Ocean, based on these field estimates of photosynthetic parameters, has been overestimated.

key words: photosynthesis-irradiance curve, quantum yield, phytoplankton, Adélie Land, Antarctica

Introduction

The Southern Ocean covers 20% of the surface of the World Ocean and plays an important role as buffer for the atmospheric greenhouse gas CO_2 . Atmospheric CO_2 is absorbed into seawater at the air-sea interface, and fixed as photosynthetic carbon by phytoplankton using light energy. Marine phytoplankton production has been estimated over several decades. Values in the Southern Ocean were once believed to be very high because of the high biomass of krill (Holm-Hansen and Mitchell, 1991). Measurements in the 1970's showed, however, that the rate of primary production in the Southern Ocean is not that high despite the generally high nutrient concentration (e.g. El-Sayed, 1970; Holm-Hansen *et al.*, 1977), and studies in the 1980's evidenced high spatial and temporal variability of primary production there (e.g. El-Sayed and Weber,

* Present address: Center for Antarctic Environment Monitoring, National Institute of Polar Research, Kaga 1-chome, Itabashi-ku, Tokyo 173-8515.
E-mail address: hirawake@nipr.ac.jp

1982; Gordon, 1988).

Several factors have been suggested that might control and reduce primary productivity in the Southern Ocean, *i.e.* temperature (Neori and Holm-Hansen, 1982), irradiance (Mitchell *et al.*, 1991), decrease of phytoplankton standing stocks by grazing of zooplankton (Smetacek *et al.*, 1990), iron (Martin *et al.*, 1990), vertical mixing of the water column (Mitchell and Holm-Hansen, 1991), and their interrelationships. Among these factors, the response of primary productivity to irradiance, characterized by the photosynthetic characteristics (photosynthesis-irradiance curve) and the quantum yield of phytoplankton, are critical to model and compute primary production from remotely sensed data. Several primary production models (*e.g.* Longhurst *et al.*, 1995) are using the field estimates of photosynthetic parameters. Thus, these parameters observed at sea directly affect the accuracy of the models.

Values for the photosynthetic characteristics have been reported for a limited region in the Southern Ocean, *i.e.* the Antarctic Peninsula region, showing low light saturation (*e.g.* Figueiras *et al.*, 1998). To better parameterize and understand the process of primary production in the Southern Ocean, and use remotely sensed information, better knowledge of the photosynthetic characteristics are needed for this whole ocean, especially the Eastern Indian Ocean sector about which we know very little.

During the 1996 cruise of the Umitaka-Marui III, Tokyo University of Fisheries, we carried out a series of measurements to determine the photosynthetic characteristics of phytoplankton in the water off Adélie Land, Antarctica. The general features of the horizontal distribution of primary productivity in the studied area have already been discussed in relation to the upper water mass structure (Chiba *et al.*, 1999). The present paper focuses on the photosynthetic parameters and quantum yield during the austral summer.

Materials and methods

Field observation and water sampling

The study was conducted off Adélie Land, Antarctica, from 20 January to 3 February 1996, during a cruise of the T/V Umitaka-Marui III (Fig. 1). Water temperature and salinity were measured with a CTD (ICTD, FSI): OCTOPUS system (OCTO-Parameter Underwater Sensor; Ishimaru *et al.*, 1984) at 27 stations. The salinity sensor was calibrated with a salinometer. Primary productivity of phytoplankton was determined at 8 stations. The ice edge was defined as 15% ice concentration, based on SSM/I images (Cavalieri *et al.*, 1997). Seawater samples for primary productivity were collected using Van Dorn bottles near the sea surface and near the bottom of the euphotic zone (irradiance=1–5% of that at sea surface). However the irradiance at 50m at Stn. C05 was 21% of that at the sea surface. Additional water samples from several depths were collected using a Rosette multi water sampler equipped with an OCTOPUS system, for determination of chlorophyll *a* concentration.

Chlorophyll *a* concentration

Between 200 and 500 ml of water were filtered onto glass fiber filters (Whatman

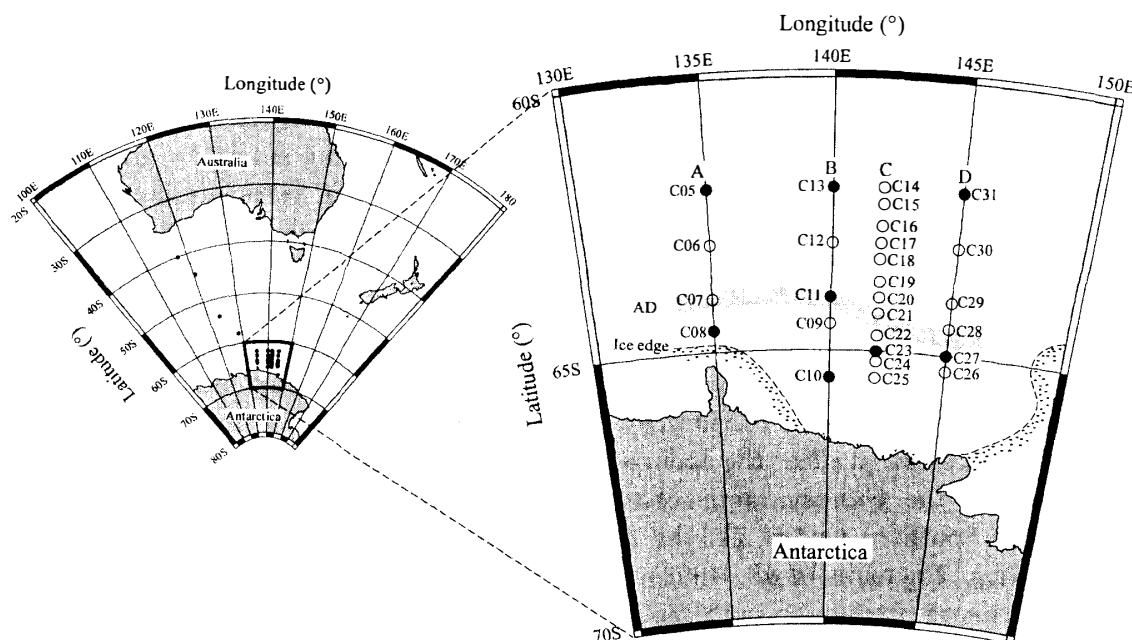


Fig. 1. Maps showing sampling stations. Solid circles: stations where the photosynthetic rate was measured. Shaded band and dashed lines: Antarctic Divergence (AD) and ice edge, respectively.

GF/F, ϕ 25 mm) for determination of phytoplankton chlorophyll *a* (chl.*a*) and pheopigments. The filters were immediately dropped in N, N-Dimethylformamide (DMF) and the pigments extracted in the dark (Suzuki and Ishimaru, 1990). Concentrations of chl.*a* were determined fluorometrically with a Turner Designs 10-005R fluorometer within a few days of extraction (Parsons *et al.*, 1984). The fluorometer was calibrated against spectrophotometric values using pure chl.*a* (SIGMA) and the concentration of the standard solution was calculated using the value of chlorophyll *a* specific absorption coefficient in DMF (Porra *et al.*, 1989).

Photosynthetically available radiation and euphotic zone

Photosynthetically available radiation (PAR) was measured with an LI-190SB air quantum sensor and an LI-192SB underwater quantum sensor (LI. COR Inc.), and recorded with an LI-1000 (LI. COR Inc.) quantum meter. Underwater PAR was measured at depths between 1 m and 20 or 30 m. The depth of the euphotic zone (Z_{eu}) was defined as the depth of 1% of sea surface PAR; it was calculated by extrapolating logarithmically the value of underwater PAR.

Photosynthetic activity

The photosynthetic activity of phytoplankton was determined using the stable ^{13}C isotope method (Hama *et al.*, 1983). Water samples were transferred into 1000 ml clear polycarbonate bottles. After adding $\text{NaH}^{13}\text{CO}_3$ (approximately 10% of the total carbonate, ISOTECH Inc.), the samples from the two sampling depths were incubated during 4 hours in a water bath with controlled temperature corresponding to that at the sea surface, under natural light (full sunlight, 46, 21, 11, 6% and, dark). After

incubation, the water samples were filtered through glass fiber filters (Whatman GF/F, ϕ 47 mm, precombusted at 450°C for 4 hours). The isotope ratios of ^{12}C and ^{13}C were determined by infrared absorption spectrometry (Sato *et al.*, 1985) with a ^{13}C analyzer (EX-130, JASCO). The photosynthetic activity was calculated using the equation of Hama *et al.* (1983), and the rate at each depth was calculated on the basis of the observed photosynthesis-irradiance (P-E) curve fit to the model of Eilers and Peeters (1988) with a nonlinear curve-fitting minimization method. The equation takes the form:

$$P^* = I / (aI^2 + bI + c), \quad (1)$$

where P^* is photosynthetic rate, I is irradiance, and a , b , and c are the fitted parameters. Photosynthetic characteristics were calculated as follows: initial slope (α^*) = $1/c$, maximum photosynthetic rate (P^*_{\max}) = $[b + 2(ac)^{0.5}]^{-1}$, and intensity of onset of light saturation (I_k) = P^*_{\max} / α^* . The irradiance, I , was the mean value to which the sample was exposed during the incubation period (Table 1).

Absorption coefficient

The absorption coefficient of particles was determined using the quantitative filter technique (Mitchell and Kiefer, 1984). The optical density, $\text{OD}(\lambda)$ between 400 and 750 nm was measured with a spectrophotometer (UV365, Shimadzu) equipped with an integrating sphere. The absorption coefficient was calculated from $\text{OD}(\lambda)$ as described by Mitchell (1990). The absorption coefficient of phytoplankton, $a_{\text{ph}}(\lambda)$, was determined by the methanol extraction method of Kishino *et al.* (1985). The chl.*a* specific absorption coefficient, $a^*_{\text{ph}}(\lambda)$, was derived by dividing $a_{\text{ph}}(\lambda)$ by the chl.*a* concentration; its units are $\text{m}^2 (\text{mg chl.}a)^{-1}$.

Maximum quantum yield

The maximum quantum yield, ϕ_m ($\text{mol C} (\text{mol photons})^{-1}$), was computed as follows:

$$\phi_m = 0.02315 \frac{\alpha^*}{\bar{a}^*_{\text{ph}}}, \quad (2)$$

where the constant (0.02315) converts grams to moles and hours to seconds. \bar{a}^*_{ph} is the spectrally averaged chl.*a* specific absorption coefficient ($\text{m}^2 (\text{mg chl.}a)^{-1}$):

$$\bar{a}^*_{\text{ph}} = \frac{\int_{400}^{700} a^*_{\text{ph}}(\lambda) \cdot I(\lambda) \cdot d\lambda}{\int_{400}^{700} I(\lambda) \cdot d\lambda}, \quad (3)$$

where $I(\lambda)$ is spectral irradiance at the sea surface measured with a spectroradiometer MER-2020A (Biospherical Instrument Inc.).

Results and discussion

Hydrographic structure, light penetration and chlorophyll *a* distribution

The vertical distributions of isotherms and isohalines in the 0–200 m layer along four transects (A, B, C and D in Fig. 1) are shown in Figs. 2 and 3, respectively. Although it is not clear in the upper water mass structure between 0 and 200 m, Chiba *et al.* (1999) showed in the structure between 0 and 4000 m that there was upwelling of warmer ($>1.0^{\circ}\text{C}$) and higher salinity (>34.5 PSU) water at around 64.0 – 64.5°S , that formed the Antarctic Divergence (AD). The water column was well stratified between 30 and 125 m, where a mass of cold water (dichothermal layer) existed that vertically separated the water column. South of the AD, there was indication of a sinking water mass, which may be a major source of Antarctic Bottom Water (Chiba *et al.*, 1999).

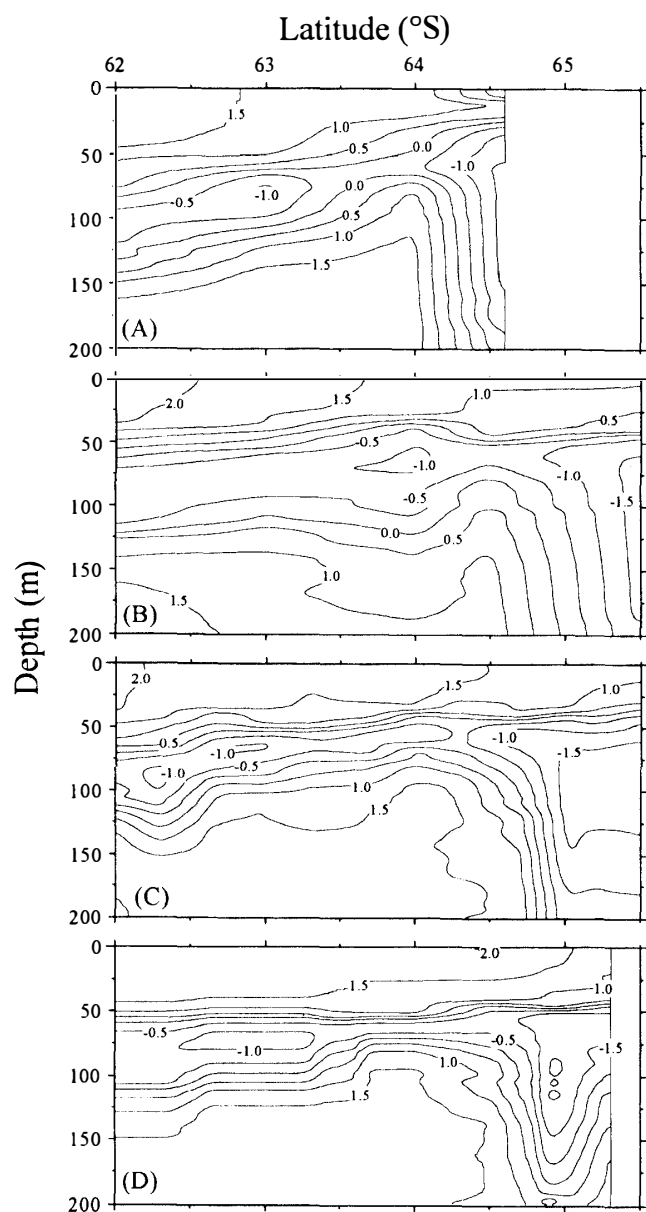


Fig. 2. Vertical distribution of isotherms ($^{\circ}\text{C}$) along transects A, B, C and D.

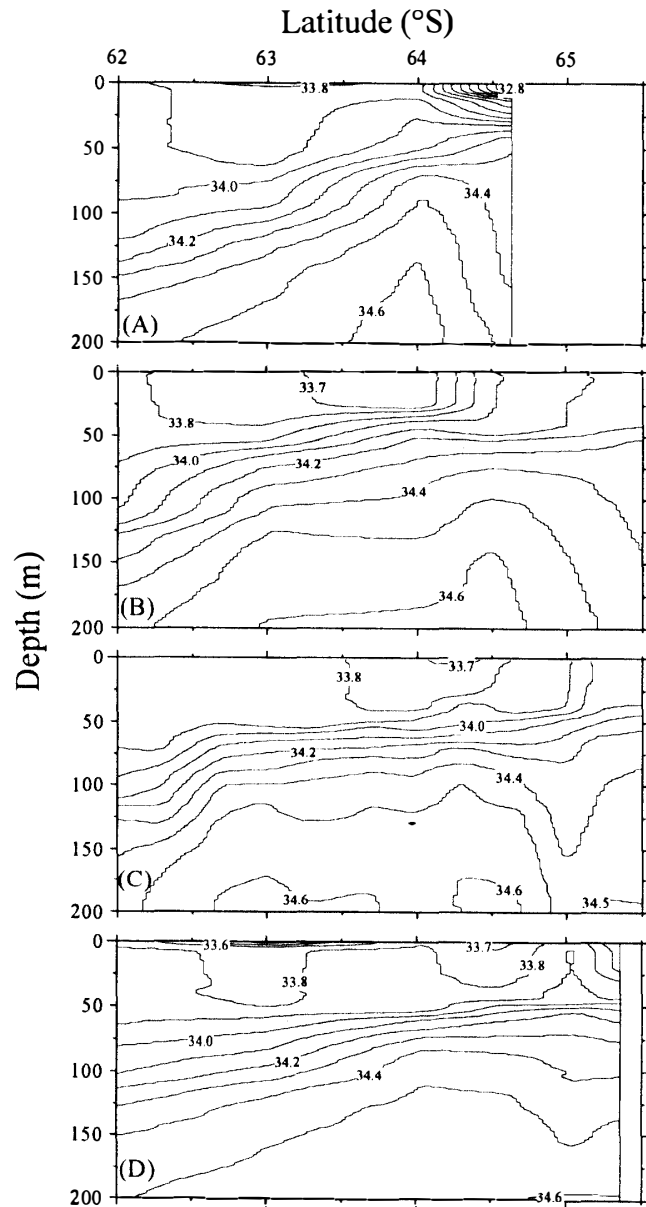


Fig. 3. Vertical distribution of isohalines (PSU) along transects A, B, C and D.

In the upper 50 m, warm ($> 1.5^{\circ}\text{C}$) and relatively high salinity (> 33.8 PSU) water extended from the northeast to southwest of the studied region.

The vertical distribution of chlorophyll *a* isopleths is shown in Fig. 4. The concentrations varied from 0.03 to 1.92 mg m^{-3} . The highest concentrations in surface waters were observed close to the AD on each transect, and near the ice edge on transect A. There was a south-north gradient with the boundary at the AD, simultaneously with an east-west gradient corresponding to that of water masses (Chiba *et al.*, 1999). The area was also characterized by a distinct subsurface chl.*a* maximum (SCM). The depth of the SCM, the mixed layer depth (MLD) and euphotic zone depth (Z_{eu}) are shown in Fig. 5. In the present study, the depth and the dispersion of the SCM were determined from the Gaussian Curve fit of Lewis *et al.* (1983) and Platt *et al.* (1988) as modified by Matsumura and Shiimoto (1993), and the MLD was defined as the upper

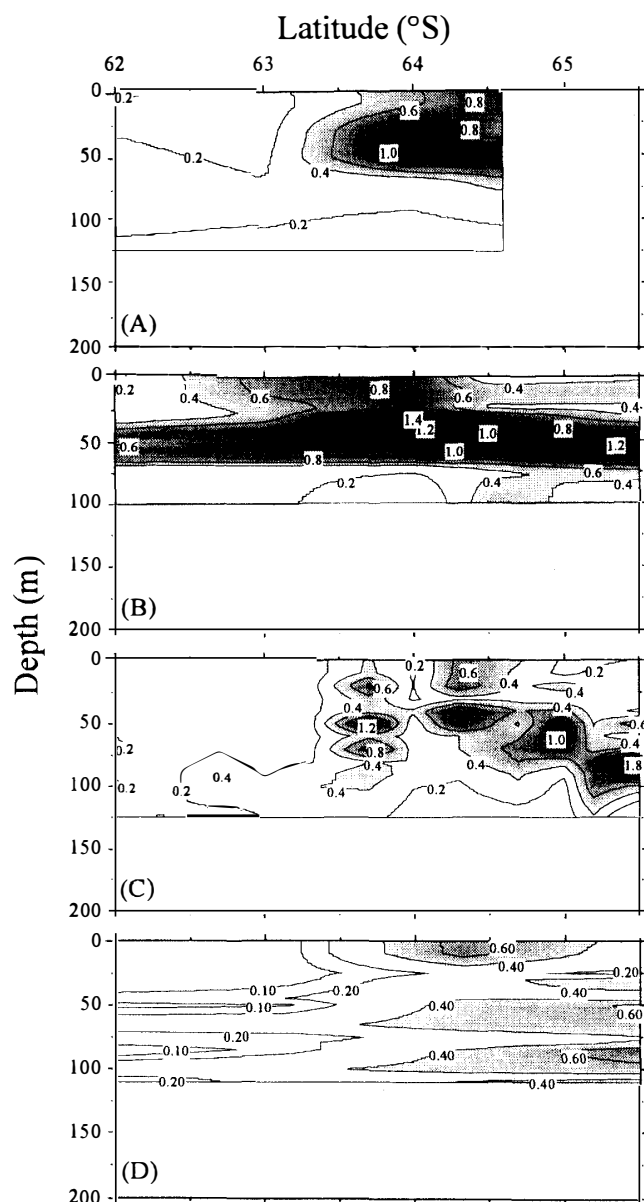


Fig. 4. Vertical distribution of chl.a concentration isopleths (mg m^{-3}) along transects A, B, C and D. Concentrations $> 0.4 \text{ mg m}^{-3}$ are shaded. The square shows the sampling depth.

depth where the temperature is 1.0°C less than that at 10 m (Rao *et al.*, 1989). The peak depth of the SCM ranged from 35 to 127 m, except at Stn. 29 (5 m). In most cases, the depth of the SCM coincided with the depth of the dichothermal water, as already reported by Yamaguchi *et al.* (1985). Furthermore, the SCM corresponded to low irradiance, as observed by El-Sayed (1988) and Holm-Hansen *et al.* (1994). Although relatively high biomass of phytoplankton in the SCM may make efficient use of input resources such as nutrients and iron from deep water, irradiance at the SCM depth was only between 1 and 12% of the incident irradiance. Therefore, phytoplankton in the SCM could not take advantage of location in the deeper water without adaptation to the low irradiance.

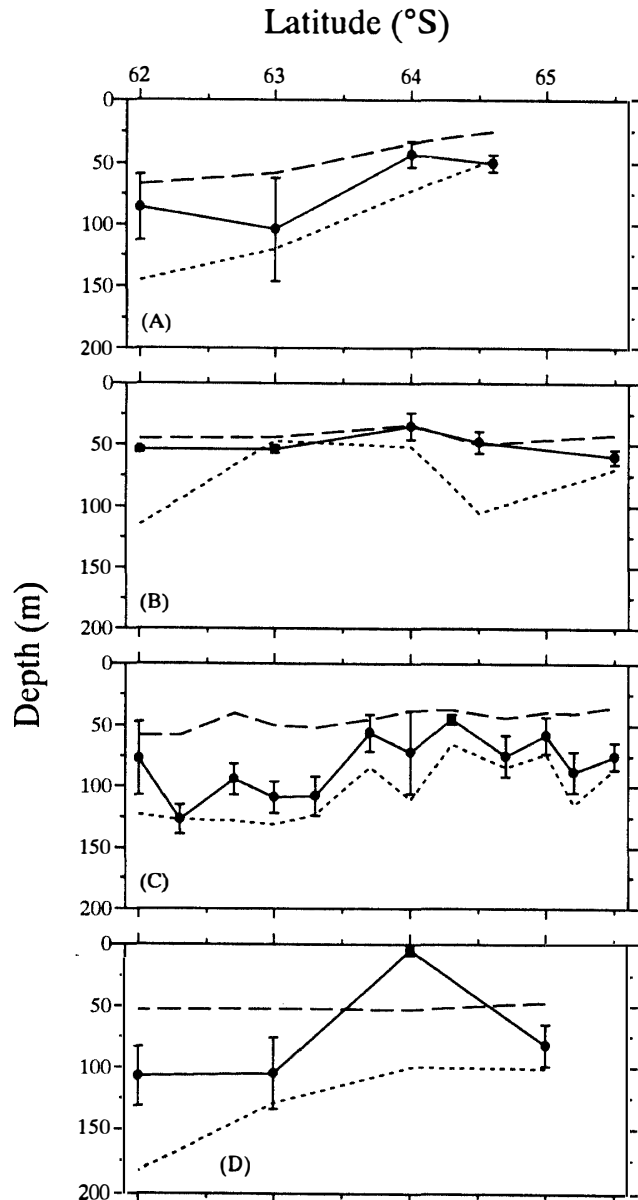


Fig. 5. Positions of the peak depth of the subsurface chl.*a* maximum (SCM; solid circle), mixed layer depth (MLD; long dashed line) and euphotic zone depth (Z_{eu} ; dashed line). The error bar on the SCM peak shows the dispersion of the SCM (\pm standard deviation).

Photosynthetic parameters

The photosynthesis–irradiance curves (P–E curves) at each station and their parameters are shown in Fig. 6 and Table 1, respectively. The initial slope (α^*) was generally high, whereas the maximum photosynthetic rate (P^*_{max}) and the onset of light saturation (I_k) were generally low, the mean values (\pm standard deviation) of α^* , P^*_{max} and I_k being $0.014 (\pm 0.013) \text{ mg C (mg chl.}a)^{-1} \text{ h}^{-1} (\mu\text{mol photons m}^{-2} \text{ s}^{-1})^{-1}$, $0.66 (\pm 0.37) \text{ mgC (mg chl.}a)^{-1} \text{ h}^{-1}$ and $76 (\pm 55) \mu\text{mol photons m}^{-2} \text{ s}^{-1}$, respectively. The relatively high values of α^* and low values of I_k indicate that phytoplankton were adapted to low irradiance. The higher values of α^* ($0.019 \text{ mgC (mg chl.}a)^{-1} \text{ h}^{-1} (\mu\text{mol photons m}^{-2} \text{ s}^{-1})^{-1}$) and lower values of I_k at depth relative to the surface suggest that the phytoplankton in the SCM had high productivity. The irradiance intensity in

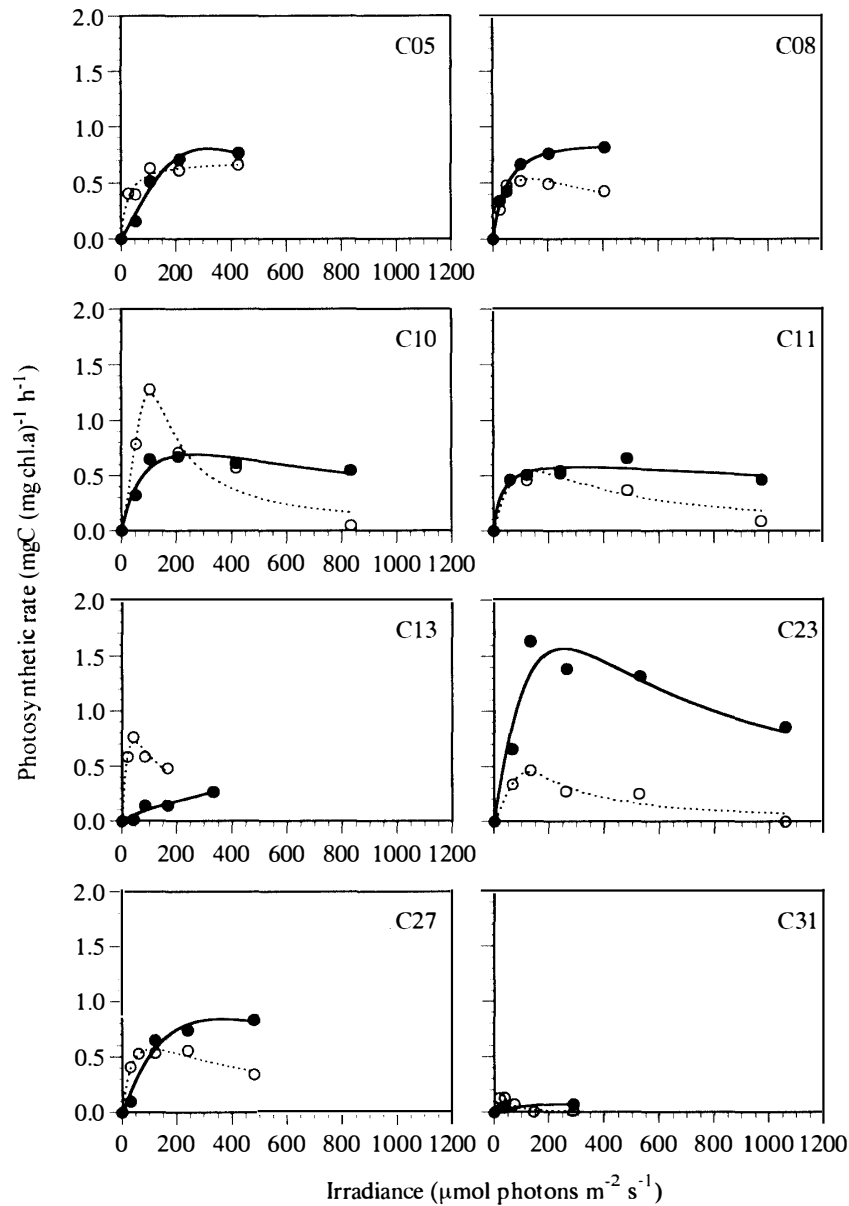


Fig. 6. P-E curves at each station. Solid circles and open circles show the data at the sea surface and near the bottom of the euphotic zone, respectively.

the SCM, computed using the average value of incident light during the cruise ($332 \mu\text{mol photons m}^{-2} \text{s}^{-1}$) was, however, lower than I_k . Thus, the phytoplankton in the SCM were not fully adapted to the low irradiance prevailing at those depths.

The highest P^*_{max} values were observed at the sea surface of Stn. C23, near the AD ($1.57 \text{ mgC (mg chl.a)}^{-1} \text{ h}^{-1}$), and at 65 m of Stn. C10, the southernmost station ($1.24 \text{ mgC (mg chl.a)}^{-1} \text{ h}^{-1}$). The lowest P^*_{max} values were observed at Stn. C31, at the surface (0.07) and at 145 m ($0.16 \text{ mgC (mg chl.a)}^{-1} \text{ h}^{-1}$). Comiso *et al.* (1990) showed enhanced phytoplankton standing stocks associated with ice retreat, from the CZCS (Coastal Zone Color Scanner) satellite imagery. We also observed the highest

Table 1. Photosynthetic characteristics of the P-E curves and maximum quantum yield.

Station	Depth (m)	I (\pm S.D.) _a	α^* _b	P^*_{\max} _c	I_k _d	ϕ^m _e
Sea surface						
C05	0	426.9 (176.5)	0.004	0.80	193	0.007
C08	0	406.3 (75.4)	0.017	0.82	47	0.031
C10	0	832.2 (162.9)	0.011	0.69	63	0.019
C11	0	974.9 (145.8)	0.020	0.58	29	0.038
C13	0	332.0 (72.5)	0.002	0.27	176	0.003
C23	0	1058.1 (138.0)	0.016	1.57	98	0.030
C27	0	478.5 (133.2)	0.007	0.84	125	0.012
C31	0	286.9 (54.8)	0.001	0.07	96	0.001
	Mean	599.5	0.010	0.71	103	0.018
	S.D.	306.2	0.007	0.45	59	0.014
Near the bottom of the euphotic zone (except C05)						
C05	50	426.9 (176.5)	0.029	0.68	23	0.043
C08	50	406.3 (75.4)	0.018	0.54	30	0.032
C10	65	832.2 (162.9)	0.011	1.24	110	0.016
C11	45	974.9 (145.8)	0.010	0.54	56	0.025
C13	75	332.0 (72.5)	0.053	0.72	14	0.092
C23	70	1058.1 (138.0)	0.005	0.45	93	0.009
C27	85	478.5 (133.2)	0.025	0.57	23	0.048
C31	145	286.9 (54.8)	0.004	0.16	41	0.007
	Mean	599.5	0.019	0.61	49	0.034
	S.D.	306.2	0.016	0.31	35	0.028
	Mean		0.014	0.66	76	0.026
	S.D.		0.013	0.37	55	0.023

^a mean irradiance to which the sample was exposed during incubation period ($\mu\text{mol photons m}^{-2} \text{ s}^{-1}$)

^b initial slope ($\text{mgC (mg chl.}a)^{-1} \text{ h}^{-1}$ ($\mu\text{mol photons m}^{-2} \text{ s}^{-1}$)⁻¹)

^c maximum photosynthetic rate ($\text{mgC (mg chl.}a)^{-1} \text{ h}^{-1}$)

^d intensity at the onset of light saturation ($\mu\text{mol photons m}^{-2} \text{ s}^{-1}$)

^e maximum quantum yield ($\text{mol C (mol photons absorbed)}^{-1}$)

surface chl.*a* concentration near the ice edge (Stn. C08), together with evidence of reduced surface water salinities (< 33.8) indicating sea-ice melting. However, the photosynthetic rate of phytoplankton was not high there.

Chiba *et al.* (1999) showed an east-west gradient in the horizontal distributions of chl.*a* concentration and dominant phytoplankton taxa, that corresponded to a water mass gradient. The photosynthetic characteristics of phytoplankton are related not only to irradiance, temperature and nutrient conditions, but also to taxonomic factors (Platt *et al.*, 1983). In the present study, only α^* at the sea surface showed a significant difference ($p = 0.0093$, *t*-test) between north and south of the AD; α^* values were lower north than south of the AD. At the surface, warm ($> 1.5^\circ\text{C}$) and relatively high

Table 2. Historical photosynthesis-irradiance curve parameters in the Southern Ocean.

Source	Date	Area	α^*	P^*_{\max}	I_k
Jacques (1983)	Mar. 1977/1980	Indian Ocean Sector (South of the Antarctic Convergence, West of Kerguelen Is.)	0.005–0.034	0.5–5.2	13.8–46.0
		Indian Ocean Sector (Continental shelf near Kerguelen Is.)	0.017–0.023	1.0–2.0	46.0–119.6
Yamaguchi <i>et al.</i> (1985)	Jan./Feb. 1984	Off Adélie Land	–	1.01, 0.73	–
Sakshaug and Holm-Hansen (1986)	Feb./Mar. 1981	Scotia/Weddell Seas	0.0094–0.049	0.75–4.4	38–190
Tilzer <i>et al.</i> (1986)	Oct./Nov. 1983	Scotia Sea and Bransfield Strait (South of PF)	0.0074	1.92	69
Holm-Hansen and Mitchell (1991)	Dec. 1986– Mar. 1987	Bransfield Strait	0.06	1.1	18
Cota <i>et al.</i> (1992)	Jun./Aug. 1988	Weddell-Scotia Sea (MIZ)	0.056	0.23	9.8
Figueiras <i>et al.</i> (1994)	Dec. 1988– Jan. 1989	Weddell, Elephant Is.– South Orkney Is.	0.058	1.94	35
Boyd <i>et al.</i> (1995)	Nov./Dec. 1992	Bellingshausen Sea	0.001–0.048	0.09–1.07	<20–90
Figueiras <i>et al.</i> (1998)	Jan. 1994	Bransfield–Bellingshausen	0.019	1.60	84
		Bransfield–Weddell	0.017	1.41	88
		Ice-Edge (Bransfield Strait)	0.022	1.63	82
		Weddell-Scotia Confluence	0.007	0.79	100
Moline <i>et al.</i> (1998)	Spring / Summer 1991–1994	Antarctic Peninsula shelf	0.043	2.99	78.44
This study	Jan./Feb. 1996	Off Adélie Land	0.014	0.66	76

salinity (>33.8 PSU) water extended from the northeast to southwest of the studied region (Figs. 2 and 3). Decline of photo-chemical response of phytoplankton in this water mass is suggested. However, there were no significant spatial differences in the photosynthetic parameters in the studied area. This may be due to a combination of several factors such as water mass structure, penetration of irradiance and dominant phytoplankton taxa among stations.

Historical photosynthetic parameters in the Southern Ocean are summarized in Table 2. Most of these data were collected from waters around the Antarctic Peninsula such as Bransfield Strait and the Weddell-Scotia Seas. Values of α^* and I_k in the present study are in the range of those previously published, but P^*_{\max} is lower than values in waters around the Antarctic Peninsula during the austral summer that frequently exceed $1.5 \text{ mgC (mg chl.}a)^{-1} \text{ h}^{-1}$. Although high P^*_{\max} was reported by Jacques (1983) in water near Kerguelen Island, P^*_{\max} in the water off Adélie Land is generally lower than in water near the Antarctic Peninsula.

The duration of incubation in the present study (4 hours) is likely to be long. Because of the rapid and large changes in ambient light to which the samples are exposed, phytoplankton have a remarkable capability for photoacclimation (Dubinsky *et al.*, 1995). The acclimation of the photosynthetic apparatus during incubation may cause P-E curve variability (Sakshaug *et al.*, 1997). Moisan and Mitchell (1999) suggested that photophysiological acclimation may contribute to the ecological success

of a prymnesiophyte, *Pheochystis antarctica*, in polar regions. Since the results in this study cannot explain the photoacclimation during the incubation period, further studies of the acclimation of phytoplankton in polar regions with various durations of incubation are required to reveal the relationship between photosynthesis and the mixing rate of the water column.

Quantum yield

The values of maximum quantum yield, ϕ_m , are shown in Table 1. These varied widely, from 0.001 to 0.038 mol C (mol photons absorbed)⁻¹ for surface water and 0.007 to 0.092 mol C (mol photons absorbed)⁻¹ near the bottom of the euphotic zone. At the surface, values >0.03 mol C (mol photons absorbed)⁻¹ were observed at Stns. C 08, C11 and C23, near the AD. Near the bottom of the euphotic zone, values >0.04 mol C (mol photons absorbed)⁻¹ occurred at Stns. C05, C13 and C27, near the AD (or the northernmost stations).

Our ϕ_m , except for the lowest (<0.01), were in the range of values from previous field studies in temperate waters (e.g. Kishino *et al.*, 1986). Smith *et al.* (1996) computed ϕ_m values in the Antarctic Peninsula region from historical α^* data. Although the computation method was slightly different from that in the present study, our values are generally within the range of these values (0.002–0.094 mol C (mol photons absorbed)⁻¹).

Conclusions

Off Adélie Land, Antarctica, the maximum photosynthetic rate of phytoplankton was lower than values in the waters near the Antarctic Peninsula, but maximum quantum yield was in the same range as in waters near the Antarctic Peninsula. Therefore, the difference of photosynthetic rate between the two regions cannot be explained by the difference in the activity of the components of electron transfer. The present study was carried out in the declining period of the Antarctic phytoplankton (El-Sayed, 1984) as well as the period studied by Yamaguchi *et al.* (1985). Studies near the Antarctic Peninsula have generally focused on the period of phytoplankton bloom during December and January. It is suggested that the difference of the period was one of the factors in the difference of P^*_{max} between the two regions.

Some primary production models on the global/regional scale using satellite data have been developed recently (e.g. Platt and Sathyendranath, 1988; Morel, 1991; Platt *et al.*, 1991; Longhurst *et al.*, 1995; Antoine *et al.*, 1995; Behrenfeld and Falkowski, 1997; Arrigo *et al.*, 1998). Some of these models use the P-E parameters and quantum yield observed at sea. Such field data for the Southern Ocean have so far been concentrated in the region near the Antarctic Peninsula. Thus, primary productivity for the whole of the Southern Ocean based on these field estimates of photosynthetic parameters is likely overestimated.

Acknowledgments

We thank the officers and crew of the T/V Umitaka-maru III, Tokyo University of

Fisheries, for their cooperation during the cruise. The research was financially supported by the Showa Shell Sekiyu Foundation for Promoting Environmental Research, and the Fund for Research in Commemoration of the Centennial Anniversary of Tokyo University of Fisheries.

References

- Antoine, D., Morel, A. and André, J.M. (1995): Algal pigment distribution and primary production in the eastern Mediterranean as derived from coastal zone color scanner observations. *J. Geophys. Res.*, **100**, 16193–16209.
- Arrigo, K.R., Worthen, D., Schnell, A. and Lizotte, M.P. (1998): Primary production in Southern Ocean waters. *J. Geophys. Res.*, **103**, 15587–15600.
- Behrenfeld, M.J. and Falkowski, P.G. (1997): Photosynthetic rates derived from satellite-based chlorophyll concentration. *Limnol. Oceanogr.*, **42**, 1–20.
- Boyd, P.W., Robinson, C., Savidge, G. and Williams, P.J.leB. (1995): Water column and sea-ice primary production during Austral spring in the Bellingshausen Sea. *Deep-Sea Res. II*, **42**, 1177–1200.
- Cavalieri, D.J., Parkinson, C.L., Gloersen, P. and Zwally, H.J. (1997): Arctic and Antarctic sea ice concentrations from multichannel passive-microwave satellite data sets: October 1978 to December 1996, User's Guide. NASA Technical Memorandum 104647, 17 p.
- Chiba, S., Hirawake, T., Horimoto, N., Satoh, R., Nakajima, Y., Ushio, S., Ishimaru, T. and Yamaguchi, Y. (1999): An overview of biological/oceanographic survey by the RTV Umitaka-Marui III off Adélie Land, Antarctica in January–February 1996. *Deep-Sea Res.* (in press)
- Comiso, J.C., Maynard, N.G., Smith, W.O., Jr. and Sullivan, C.W. (1990): Satellite ocean color studies of Antarctic ice edges in summer and autumn. *J. Geophys. Res.*, **95**, 9481–9496.
- Cota, G.F., Smith, W.O., Jr., Nelson, D.M., Muench, R.D. and Gordon, L.I. (1992): Nutrient and biogenic particulate distributions, primary productivity and nitrogen uptake in the Weddell-Scotia Sea marginal ice zone during winter. *J. Mar. Res.*, **50**, 155–181.
- Dubinsky, Z., Matsukawa, R. and Karube, I. (1995): Photobiological aspects of algal mass culture. *J. Mar. Biotechnol.*, **2**, 61–65.
- Eilers, P.H.C. and Peeters, J.C.H. (1988): A model for relationship between light intensity and the rate of photosynthesis in phytoplankton. *Ecol. Model.*, **42**, 199–215.
- El-Sayed, S.Z. (1970): On the productivity of the Southern Ocean (Atlantic and Pacific Sectors). *Antarctic Ecology I*, ed. by M.W. Holdgate. London, Academic Press, 119–135.
- El-Sayed, S. (1984): Productivity of the Antarctic waters—A reappraisal. *Marine Phytoplankton and Productivity*, ed. by O. Holm-Hansen *et al.* Berlin, Springer-Verlag, 19–34.
- El-Sayed, S.Z. (1988): Seasonal and interannual variabilities in Antarctic phytoplankton with reference to krill distribution. *Antarctic Ocean and Resources Variability*, ed. by D. Sahrhage. Berlin, Springer-Verlag, 101–119.
- El-Sayed, S.Z. and Weber, L.H. (1982): Spatial and temporal variations in phytoplankton biomass and primary productivity in the Southwest Atlantic and the Scotia Sea. *Polar Biol.*, **1**, 83–90.
- Figueiras, F.G., Perez, F.F., Pazos, Y. and Rios, A.F. (1994): Light and productivity of Antarctic phytoplankton during austral summer in an ice edge region in the Weddell-Scotia Sea. *J. Plankton Res.*, **16**, 233–253.
- Figueiras, F.G., Estrada, M., Lopez, O. and Arbones, B. (1998): Photosynthetic parameters and primary production in the Bransfield Strait: relationships with mesoscale hydrographic structures. *J. Mar. Sys.*, **17**, 129–141.
- Gordon, A.L. (1988): Spatial and temporal variability within the Southern Ocean. *Antarctic Ocean and Resources Variability*, ed. by D. Sahrhage. Berlin, Springer-Verlag, 41–56.
- Hama, T., Miyazaki, T., Ogawa, Y., Iwakuma, T., Takahashi, M., Otsuki, A. and Ichimura, S. (1983): Measurement of photosynthetic production of a marine phytoplankton population using a stable ¹³C isotope. *Mar. Biol.*, **73**, 31–36.
- Holm-Hansen, O. and Mitchell, B.G. (1991): Spatial and temporal distribution of phytoplankton and primary production in the western Bransfield Strait region. *Deep-Sea Res.*, **38**, 961–980.

- Holm-Hansen, O., El-Sayed, S.Z., Franceschini, G.A. and Cuhel, R.L. (1977): Primary production and the factors controlling phytoplankton growth in the Southern Ocean. Adaptations within Antarctic Ecosystems, ed. by G.A. Llano. Houston, Gulf Publ. Co., 11–50.
- Holm-Hansen, O., Amos, A.F., Silva, N.S., Villafane, V. and Helbing, E.W. (1994): *In situ* evidence for a nutrient limitation of phytoplankton growth in pelagic Antarctic waters. *Antarct. Sci.*, **6**, 315–324.
- Ishimaru, T., Otake, H., Saino, T., Hasumoto, H. and Nakai, T. (1984): OCTOPUS, an octo parameter underwater sensor, for use in biological oceanography studies. *J. Oceanogr. Soc. Jpn.*, **40**, 207–212.
- Jacques, G. (1983): Some ecophysiological aspects of the Antarctic phytoplankton. *Polar Biol.*, **2**, 27–33.
- Kishino, M., Takahashi, M., Okami, N. and Ichimura, S. (1985): Estimation of the spectral absorption coefficients of phytoplankton in the sea. *Bull. Mar. Sci.*, **37**, 634–642.
- Kishino, M., Okami, N., Takahashi, M. and Ichimura, S. (1986): Light utilization efficiency and quantum yield of phytoplankton in a thermally stratified sea. *Limnol. Oceanogr.*, **31**, 557–566.
- Lewis, M.R., Cullen, J.J. and Platt, T. (1983): Phytoplankton and thermal structure in the upper ocean: Consequences of nonuniformity in chlorophyll profile. *J. Geophys. Res.*, **88**, 2565–2570.
- Longhurst, A., Sathyendranath, S., Platt, T. and Caverhill, C. (1995): An estimate of global primary production in the ocean from satellite radiometer data. *J. Plankton Res.*, **17**, 1245–1271.
- Martin, J.H., Gordon, R.M. and Fitzwater, S.E. (1990): Iron in Antarctic waters. *Nature*, **345**, 156–158.
- Matsumura, S. and Shiimoto, A. (1993): Vertical distribution of primary productivity function Φ (II)—for the estimation of primary productivity using by satellite remote sensing—. *Bull. Nat. Res. Inst. Far Seas Fish.*, **30**, 227–270. (in Japanese with English abstract)
- Mitchell, B.G. (1990): Algorithms for determining the absorption coefficient of aquatic particulates using the quantitative filter technique (QFT). *Ocean Optics X*, ed. by R.W. Spinrad. SPIE 1302, 137–148.
- Mitchell, B.G. and Kiefer, D.A. (1984): Determination of absorption and fluorescence excitation spectra for phytoplankton. *Marine Phytoplankton and Productivity*, ed. by O. Holm-Hansen *et al.* Berlin, Springer-Verlag, 157–169.
- Mitchell, B.G. and Holm-Hansen, O. (1991): Observations and modeling of the Antarctic phytoplankton crop in relation to mixing depth. *Deep-Sea Res.*, **38**, 981–1007.
- Mitchell, B.G., Brody, E.A., Holm-Hansen, O., McClain, C. and Bishop, J. (1991): Light limitation of phytoplankton biomass and macronutrient utilization in the Southern Ocean. *Limnol. Oceanogr.*, **36**, 1662–1677.
- Moisan, T.A. and Mitchell, B.G. (1999): Photophysiological acclimation of *Phaeocystis antarctica* Karsten under light limitation. *Limnol. Oceanogr.*, **44**, 247–258.
- Moline, M.A., Schofield, O. and Boucher, N.P. (1998): Photosynthetic parameters and empirical modelling of primary production: a case study on the Antarctic Peninsula shelf. *Antarct. Sci.*, **10**, 45–54.
- Morel, A. (1991): Light and marine photosynthesis: a spectral model with geochemical and climatological implications. *Prog. Oceanogr.*, **26**, 263–306.
- Neori, A. and Holm-Hansen, O. (1982): Effect of temperature on rate of photosynthesis in Antarctic phytoplankton. *Polar Biol.*, **1**, 33–38.
- Parsons, T.R., Maita, Y. and Lalli, C.M. (1984): A manual of Chemical and Biological Methods for Seawater Analysis. New York, Pergamon Press, 173 p.
- Platt, T. and Sathyendranath, S. (1988): Oceanic primary production: Estimation by remote sensing at local and Regional Scales. *Science*, **241**, 1613–1620.
- Platt, T., Subba Rao, D.V. and Irwin, B. (1983): Photosynthesis of picoplankton in the oligotrophic ocean. *Nature*, **300**, 702–704.
- Platt, T., Sathyendranath, S., Caverhill, C.M. and Lewis, M.R. (1988): Ocean primary production and available light: further algorithms for remote sensing. *Deep-Sea Res.*, **35**, 855–897.
- Platt, T., Caverhill, C. and Sathyendranath, S. (1991): Basin-scale estimates of oceanic primary production by remote sensing: The North Atlantic. *J. Geophys. Res.*, **96**, 15147–15159.
- Porra, R.J., Thompson, W.A. and Kriedemann, P.E. (1989): Determination of accurate extinction coefficients and simultaneous equations for assaying chlorophylls *a* and *b* extracted with four different solvents: verification of the concentration of chlorophyll standards by atomic absorption spectroscopy. *Biochim. Biophys. Acta*, **975**, 384–394.
- Rao, R.R., Molinari, R.L. and Festa, J.F. (1989): Evolution of the climatological near-surface thermal

- structure of the tropical Indian ocean. 1. Description of mean monthly mixed layer depth, and sea surface temperature, surface current, and surface meteorological fields. *J. Geophys. Res.*, **94**, 10801–10815.
- Sakshaug, E. and Holm-Hansen, O. (1986): Photoadaptation in Antarctic phytoplankton: variations in growth rate, chemical composition and *P* versus *I* curves. *J. Plankton Res.*, **8**, 459–473.
- Sakshaug, E., Bricaud, A., Dandonneau, Y., Falkowski, P.G., Kiefer, D.A., Legendre, L., Morel, A., Parslow, J. and Takahashi, M. (1997): Parameters of photosynthesis: definitions, theory and interpretation of results. *J. Plankton Res.*, **19**, 1637–1670.
- Satoh, H., Yamaguchi, Y., Kokubun, N. and Aruga, Y. (1985): Application of infrared absorption spectrometry for measuring photosynthetic production of phytoplankton by the stable ¹³C isotope method. *La Mer*, **23**, 171–176.
- Smetacek, V., Scharek, R. and Nothig, E.M. (1990): Seasonal and regional variation in the pelagial and its relationship to the life history cycle of krill. *Antarctic Ecosystems. Ecological Change and Conservation*, ed. by K.R. Kerry and G. Hempel. Berlin, Springer-Verlag, 103–114.
- Smith, R.C., Dierssen, H.M. and Vernet, M. (1996): Phytoplankton biomass and productivity in the Western Antarctic Peninsula region. *Antarct. Res. Ser.*, **70**, 333–356.
- Suzuki, R. and Ishimaru, T. (1990): An improved method for the determination of phytoplankton chlorophyll using N,N-Dimethylformamide. *J. Oceanogr. Soc. Jpn.*, **46**, 190–194.
- Tilzer, M.M., Elbrachter, M., Gieskes, W.W. and Beese, B. (1986): Light-temperature interactions in the control of photosynthesis in Antarctic phytoplankton. *Polar Biol.*, **5**, 105–111.
- Yamaguchi, Y., Kosaki, S. and Aruga, Y. (1985): Primary productivity in the Antarctic Ocean during the austral summer of 1983/84. *Trans. Tokyo Univ. Fish.*, **6**, 67–84.

(Received August 3, 1999; Revised manuscript accepted September 8, 1999)